



1 **METHODS FOR DETERMINING THERAPEUTIC BENEFICIAL RESONANT**
2 **FREQUENCIES**

3
4 **CROSS REFERENCE TO RELATED APPLICATIONS**

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6 This application claims priority to applicant's co-pending application having U.S. Serial
7
8 No. 60/181,460, filed February 10, 2000.

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11 **FIELD OF THE INVENTION**

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13 The present invention relates to methods for determining resonant frequencies having
14 therapeutic beneficial uses in a variety of settings. In particular, the present invention provides
15 methods for efficiently determining therapeutic useful resonant frequencies for to influence
16 ~~biological nucleic acids, in particular~~ complete genomes ~~genomes of pathogens~~ or partial
17 genomic materials, ~~composed of DNA or RNA~~ for use in various ~~which may be present in a~~
18 ~~variety of surrounding media~~ having different refractivities. ~~that may have velocities of~~
19 ~~propagation of electromagnetic waves different from that of air.~~

20
21
22 **BACKGROUND OF THE INVENTION**

23
24 Resonant frequency therapy (RFT) is a non-invasive treatment that has been reported to
25 offer significant relief to sufferers of a variety of ailments and medical conditions. The use of
26 RFT for human and animal therapeutic purposes began in the early 1900's, and experienced
27 accelerated development through the research of Royal Rife and his associates in the 1930's and
28 afterward.

29 Using new microscope technology he developed, ~~Rife observed that specific disease-~~
30 ~~causing microorganisms each responded to a definite and distinct frequency.~~ Rife discovered that
31 plasma waves could be used to transmit radio and audio frequencies, which were tuned to the

1 frequencies of specific microorganisms, and that each microorganism ~~repeatedly~~ responded to its
2 unique ~~frequency or frequencies emitted from a plasma emission device~~. For example, Rife
3 found that staphylococcus, streptococcus, microorganisms associated with tuberculosis, typhoid,
4 and leprosy, as well as cancer particles, and other disease-causing agents succumbed when
5 exposed to certain frequencies peculiar to each organism or particle. See, Siedel, R.E., and M.E.
6 Winter, The New Microscopes, Smithsonian Annual Report 1944, pp. 193-200.

7 Using the principles of Rife's discoveries, various researchers developed devices for
8 emitting frequencies designed to treat a range of diseases and conditions. For example, Dr.
9 Abraham Ginsberg used an apparatus which produced intermittent bursts of high energy in the
10 short wave spectrum. Ginsberg's modality was found to stimulate the reticuloendothelial system
11 without undesirably heating tissue. Using his device, Ginsberg reported successfully treating
12 patients with various clinical conditions, including chronic Staphylococcus infections, acute
13 inflammatory middle ear, chronic ulcerative colitis, bronchitis, rheumatoid arthritis, gout, flu,
14 and thrombophlebitis, among others. See, Cominole, B., Clinical Impressions and Speculations
15 on the Use of High-Frequency Pulsed Energy, The Dr. Abraham J. Ginsberg Foundation for
16 Medical Research Symposium, June 29, 1959.

17 Research utilizing resonant frequencies and therapeutic modalities implementing such
18 frequencies have proliferated over the past ~~ten~~ fifteen years. A recent example of the use of
19 resonant frequency therapy is the Christchurch Resonant Frequency Therapy Centre in Dunedin,
20 New Zealand. While the Centre emphasizes that resonant frequency therapy is not intended to
21 replace treatment regimens and medication prescribed by physicians, it does report successful
22 treatment of a range of clinical conditions, including arthritis, tinnitus, blood pressure, cataracts,
23 headaches, shingles, and psoriasis. Arthritis patients report particular success with pain

1 reduction and greater mobility. See The Christchurch Press, Frequency Therapy Offers Relief,
2 Independent Newspapers Limited, Oct. 28, 1999.

3 Thus, the use of ~~electric fields and/or magnetic fields, delivered with audio~~ audio range,
4 radio radio range, and light visible range frequency waves to inhibit microbial growth and to
5 treat diseases and affected tissue is well known in the art. Effective therapeutic beneficial
6 resonant frequencies have been identified through various means. Trial ~~Numerous trial~~ and error
7 approaches with resonant frequencies have been used ~~through the course of many years~~ to obtain
8 therapeutic beneficial responses. Devices for applying electromagnetic energy to living tissue
9 are disclosed, for example, in U.S. Patent No. 3,876,373, U.S. Patent No. 4,524,079, and U.S.
10 Patent No. 5,091,152. Effective resonant frequencies have also been identified through the use
11 of frequency scanning with electronic devices capable ~~claiming the capability~~ of detecting a
12 frequency response from a bacterial, viral, and/or tissue sample. Such devices for detecting
13 frequency response are disclosed, for example, in U.S. Patent No. 5,552,274, U.S. Patent No.
14 5,981,182, and U.S. Patent No. 6,004,257. Thus, there exists a need for a more efficient and
15 accurate ~~method~~ methodology than trial and error, to determine therapeutic resonant frequencies
16 ~~useful against~~ for specific target materials, such as microorganisms.

17 Therapeutic resonant frequencies may be used to inhibit, or debilitate, and/or ~~or~~
18 ~~conversely to~~ stimulate a biophysical event. The efficacy of such frequencies, whether for
19 stimulation or for debilitation, depends to some extent on the type of frequency delivery system
20 used, including variables such as power levels, waveform, harmonic content of the wave, and
21 other factors. Once therapeutic beneficial resonant frequencies are determined, the user must
22 choose which devices and delivery systems are most effectively used in conjunction with those

1 frequencies. To increase ~~general~~ efficacy, an easier, quicker, and more accurate way of
2 determining therapeutic resonant frequencies is needed.

3 Despite both historical and increasing recent interest in use of resonant frequency
4 therapy, mechanism(s) of action underlying the use of known therapeutic resonant ~~accepted~~
5 ~~beneficial~~ frequencies is not fully understood. While it is ~~generally~~ recognized that some type of
6 resonance phenomenon debilitates or destroys ~~underlies the debilitation or destruction of~~
7 microorganisms, the biophysical and/or biochemical mechanism(s) associated with use of
8 specific resonant frequencies and that lead to microbial inhibition are not completely known.

9 Before now, there has never existed a methodology that links effective therapeutic
10 resonant frequencies to a biophysical or biochemical event, process, or structure. The electronic
11 scanning devices and methods currently commercially available provide no explanation or
12 insight regarding which physical structure or process is influenced by the frequencies used.

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14
15 One entire paragraph from the previously submitted specification should be deleted at this
16 location. It is located beginning at page 4 line 15 of that document. The text begins “In PCT
17 patent application WO 8403165”, and ends “to achieve the desired biophysical event”.

18
19
20 There is a need for methodology to more readily and efficiently ~~determine wavelengths~~
21 ~~and frequencies intended to influence specific nucleic acid materials.~~ The methodology would
22 ~~provide for precise adjustments of the wavelength as required by the refractive index of a~~
23 ~~surrounding medium, and the corresponding frequency could then~~ influence genomic materials.

1 by more precisely and efficiently determining therapeutic resonant frequencies that can be easily
2 and accurately adjusted to ranges used by currently available devices. It is to these perceived
3 needs that the present invention is directed.

4 5 6 SUMMARY OF INVENTION

7
8 The present invention provides methods for determining resonant frequencies having
9 therapeutic beneficial uses outcome in a variety of settings. In particular, the present invention
10 provides methods for efficiently and accurately determining therapeutic resonant frequencies for
11 complete genomes and partial genomic materials, ~~and nucleic acids of biological origin,~~ for use
12 ~~in conjunction with~~ various media having different refractivities. ~~velocities of propagation of~~
13 ~~electromagnetic waves different than that of air.~~

14 Methods of the present invention utilize biophysical and biochemical properties of
15 genomic materials ~~nucleic acids of biological origin and their surrounding media,~~ to determine
16 therapeutic resonant wavelengths and frequencies. For example, the length of any object can be
17 considered as having a resonant frequency by virtue of correlation with a wavelength that
18 manifests itself into a ~~is presented to its innate material, or into the immediately~~ surrounding
19 ~~medium. or atmosphere. A very common example of such resonance is connected with the~~
20 ~~height of the human body. It is well accepted that certain ranges of radio frequency wavelengths~~
21 ~~that are related to human heights will cause resonance and increased absorption of energy from~~
22 ~~the wave. For that reason, those particular bands of radio wavelengths cannot be safely used for~~
23 ~~broadcasting. On that basis, Using the very same concept,~~ the length of biomolecular chains of
24 DNA and RNA can be calculated, and thus can provide wavelength-matching information
25 unique to a specific strand of genomic material. nucleic acid.

1 DNA or RNA chains are constructed in such a way that negatively-charged molecular
2 ions (the PO₄ groups) run the entire length of the molecule on the outer surface of the chain in a
3 helical fashion, causing the molecule to contain a relatively large negative charge on its surface.
4 ~~The medium surrounding these nucleic acid chains also contain a large number of positive ions~~
5 ~~(termed the “Manning cloud”), as well as polar water molecules that orient their positive side~~
6 ~~toward the negatively charged chain.~~ Thus the chain is ~~and its surrounding medium would be~~
7 highly electro-sensitive to the influences of resonant ~~external~~ oscillating electromagnetic fields.
8 Resonance is defined as the increase in amplitude of the natural oscillation or frequency of a
9 system, when exposed to a an-external periodic force whose frequency is equal or very close to
10 the natural frequency of the system. The natural oscillation of a system or part of a system ~~in~~
11 ~~time~~ is defined as its “natural resonant frequency”. ~~and is intimately linked with how the entire~~
12 ~~length of the wave travels through the system.~~ As an example, when a system, such as a strand
13 of DNA, is exposed to a frequency that presents a wavelength which is the same or very close to
14 the innate length parameter of the particular DNA strand, the motion of the externally emitted
15 wave can cause an increase in motional or electronic response of the DNA and its surrounding
16 medium, causing the DNA to resonate.

17 In radio science, the length of an antenna will largely determine how effectively the
18 antenna responds to the wavelength energy of an incoming transmission. Methods for
19 determining therapeutic resonant frequencies of the present invention utilize the principle that the
20 length of a DNA or RNA helical chain can be electromagnetically resonated in similar fashion.

21 Methods of the present invention allow precise correlations between resonant frequencies
22 and the length parameter of the genomic material ~~nucleic acid chain~~ under consideration. If a
23 resonant frequency ~~and its associated wavelength~~ is generated in air (or a vacuum) while the

1 target material ~~nucleic acid chain~~ resides in a different medium, in this invention's method a
2 refractive adjustment is made to insure that the wavelength traveling from the air or vacuum
3 medium transforms to the ~~wavelength~~ length of the target material in the surrounding medium.
4 By accounting for an appropriate ~~making use of the~~ electromagnetic refractive index for
5 ~~associated with the specific~~ surrounding medium, such as water or tissue, methods of the present
6 invention provide the advantage of determining a resonant frequency that would be more closely
7 related to ~~the innate length parameter~~ the length of the genomic material and ~~the~~ its natural
8 resonant frequency, and thus would be more appropriate, or therapeutic, for the genomic material
9 ~~nucleic acid chain~~ in that specific medium.

10 The natural electromagnetic resonant frequencies for ~~innate length of most DNA or RNA~~
11 ~~genomes if considered to be a wavelength, would most often~~ fall for the most part in the infrared
12 region of the electromagnetic (EM) spectrum. The natural resonant frequencies ~~similar~~
13 ~~associated wavelengths for very small genomes, genes and smaller portions of nucleic acid~~
14 ~~chains would~~ DNA or RNA appear in the near infrared, visible, and near ultraviolet regions of
15 the spectrum. For many currently available frequency-emitting ~~or wavelength-generating~~
16 devices, the natural resonant frequencies ~~emissions capable of such high spectrum wavelengths~~
17 such as those associated with genomic ~~nucleic acid~~ material are not achievable due to the
18 technical limitations ~~or in some cases the price~~ of the device. Indeed, particular devices often are
19 capable of generating frequencies in only narrow ~~electromagnetic frequency~~ ranges. To
20 overcome such limitations, methods of the present invention adjust resonant frequencies upward
21 or downward. For example, to determine an appropriate lower range frequency in accordance
22 with the present invention, the therapeutic resonant frequency is divided by the number 2, as
23 many times as necessary, until a frequency in the frequency-generating range of a device is

1 reached. The ~~actual~~ power of 2 by which a therapeutic ~~an original~~ resonant frequency is
2 factored ~~divided~~, will depend on the range of the electromagnetic spectrum within which a
3 frequency delivery device operates.

4 In music, a similar adjustment would be termed moving to a higher or lower octave.
5 Moving to a higher octave would in effect cut the wavelength in half, while moving to a lower
6 octave would double the wavelength. In accordance with methods of the present invention,
7 therapeutic resonant frequencies of genomic material ~~associated with nucleic acid chains are~~
8 ~~translated, or~~ “shifted by octaves,” to a lower octave in the electromagnetic spectrum, by
9 dividing the therapeutic resonant frequency by ~~some~~ an appropriate power of the number 2. The
10 lower octave of a therapeutic resonant frequency, while having a much longer wavelength, will
11 resonate with the first therapeutic ~~original high octave~~ resonant frequency, just as musical
12 octaves resonate with and amplify each other, but only when the octave ~~translation~~ shift is exact.
13 ~~Thus, to be effective, a lower octave resonant frequency must have a precise power of 2~~
14 ~~correlation with the original high octave resonant frequency of the target material. Likewise, if~~
15 ~~an octave related resonant frequency is chosen which is higher than the original resonant~~
16 ~~frequency, the higher octave resonant frequency would be accurately determined by multiplying~~
17 ~~the original one by a precise power of 2.~~

18 The present invention comprises methods for determining therapeutic resonant
19 frequencies of electromagnetic radiation ~~emission~~ for influencing a target genomic material
20 ~~nucleic acid chain~~, where the genomic material ~~chain~~ is surrounded by a medium. Embodiments
21 of these methods include the following steps: (1) determining a velocity of electromagnetic
22 radiation ~~emission~~ through the medium surrounding the genomic ~~nucleic acid~~ material; (2)
23 determining the length of the genomic ~~nucleic acid~~ material; (3) determining a first resonant

1 frequency of the genomic nucleic acid material in one electromagnetic frequency range by
2 dividing the velocity of the electromagnetic radiation through ~~emission associated with the~~
3 surrounding medium by the length of the genomic material ~~nucleic acid chain~~; (4) dividing or
4 multiplying the first resonant frequency by a factor of a power of two to obtain at least one of a
5 ~~group of resonant frequencies~~ frequency in at least one other another electromagnetic frequency
6 range; (5) programming a frequency-emitting ~~frequency-capable emission~~ device to emit the at
7 least one of a ~~group of resonant frequencies~~ frequency in the at least one other electromagnetic
8 frequency range selected in step 4; and (6) selectively influencing the target genomic nucleic
9 ~~acid chain~~ material with the at least one of a ~~group of resonant frequencies~~ frequency in the at
10 ~~least one other~~ selected electromagnetic frequency range, when the frequency-emitting
11 ~~frequency-capable emission~~ device emits the at least one of a ~~group of resonant frequencies~~
12 frequency in the at least one other selected electromagnetic frequency range into the medium
13 surrounding the target genomic ~~nucleic acid chain~~ material.

14 Methods of the present invention further comprise determining the length ~~parameter~~ of
15 the genomic target nucleic acid chain material by determining ~~obtaining~~ the number of
16 nucleotides base pairs in a single strand of the target nucleic acid chain the genomic material (in
17 the case of single-stranded genomic material, this step would comprise determining the number
18 of bases); ~~and in the case of double-stranded molecules not including the number of nucleotide~~
19 ~~bases in the complementary strand using the known value for the average spacing between~~
20 adjacent ~~nucleotide bases~~ base pairs or bases; and multiplying the number of ~~nucleotides~~ base
21 pairs or bases in the ~~target nucleic acid chain~~ genomic material by the ~~known average spacing~~
22 ~~value between adjacent nucleotides~~ base pairs or bases. In a preferred embodiment, the
23 nucleotides base pairs or bases are spaced apart by an average spacing, which is a known value,

1 and determining the length of the genomic material ~~nucleic acid chain~~ comprises ~~obtaining~~
2 ~~determining~~ the number of ~~nucleotides~~ base pairs or bases in the genomic material ~~chain~~, and
3 then multiplying that number of ~~nucleotides~~ base pairs or bases in the genomic material ~~chain~~ by
4 the known value for the average spacing between ~~nucleotides~~ base pairs or bases. As will be
5 ~~obvious to those with minimal knowledge of the art, in the case of double stranded nucleic acid~~
6 ~~chains, the number of nucleotides included in the count should include only one side of the~~
7 ~~double chain, in order to not calculate a chain length twice as long than it actually is.~~

8 In a typical environment, genomic biological ~~nucleic acid chain~~ material exists in living,
9 or in-vivo, tissue. In methods of the present invention, the velocity of electromagnetic radiation
10 ~~emissions~~ through in-vivo tissue ~~is~~ can be determined by accounting for ~~obtaining~~ the unique
11 electrical permittivity ~~value of associated with~~ in-vivo tissue in relation to velocity, and then
12 ~~determining the in-vivo associated velocity~~ such that the velocity = $1 / \sqrt{(\epsilon \mu)}$, where ϵ is the
13 electrical permittivity of ~~the medium~~ in-vivo tissue, and μ is the magnetic permeability of the
14 medium in-vivo tissue. With Having this measurement of in-vivo velocity, a refractive index of
15 electromagnetic radiation through in-vivo tissue is determined by dividing the velocity of
16 electromagnetic radiation, or the speed of light in a vacuum, by the speed of light in in-vivo
17 tissue. Then by dividing a therapeutic resonant frequency determined for the genomic material
18 in an air medium by the refractive index for in-vivo tissue, a therapeutic resonant frequency for
19 the genomic material surrounded by in-vivo tissue is determined. ~~an initial resonant frequency~~
20 ~~relating to the target nucleic acid chain can be determined by dividing the in-vivo velocity by the~~
21 ~~length of the nucleic acid chain under consideration. This step constitutes using the physics~~
22 ~~relationship, velocity = frequency times wavelength, or in its variation, velocity divided by~~
23 ~~wavelength = frequency.~~

1 In other embodiments, methods of the present invention include multiplying therapeutic
2 resonant frequencies in ~~a the range~~ adaptable for use in frequency-emitting devices ~~used by an~~
3 ~~emission device~~ by a positive integer to determine harmonic frequencies; or dividing therapeutic
4 resonant frequencies in ~~a the range~~ adaptable for use in frequency-emitting devices ~~used by an~~
5 ~~emission device~~ by a positive integer to determine subharmonic frequencies. By programming a
6 frequency-emitting device to emit the harmonic and subharmonic frequencies, target genomic
7 ~~Nucleic acid~~ material is ~~can also be~~ selectively influenced with the therapeutic resonant
8 frequencies and the harmonic and subharmonic frequencies, harmonics or subharmonics of the
9 ~~aforementioned resonant frequency, when the frequency-emitting frequency-capable emission~~
10 device emits these ~~is programmed to emit the harmonically derived~~ frequencies into the medium
11 surrounding the target genomic material. ~~nucleic acid chain.~~

12 Features of methods for determining therapeutic resonant frequencies of the present
13 invention may be accomplished singularly, or in combination, in one or more of the
14 embodiments of the present invention. As will be appreciated by those of ordinary skill in the
15 art, the present invention has wide utility in a number of applications as illustrated by the variety
16 of features and advantages discussed below.

17 Methods of the present invention provide numerous advantages over prior efforts to
18 identify therapeutic resonant frequencies. For example, the present invention advantageously
19 provides methods for determining resonant frequencies effective for stimulation and/or
20 debilitation of specific types of DNA and/or RNA genomes, genes and gene sections. ~~and nucleic~~
21 ~~acid chains.~~

1 Another advantage of the methods of the present invention is that they provide means for
2 readily and efficiently determining therapeutic resonant frequencies using widely ~~publicly~~
3 available data.

4 Another advantage is that the present invention provides methods for readily and
5 efficiently predicting resonant frequencies that can be used therapeutically or beneficially in a
6 variety of ~~settings~~ circumstances ~~surrounding microbiological and biochemical events, including~~
7 related to treatment of various human and animal diseases and conditions, ~~agriculture-related~~
8 ~~diseases, pathogen contamination of water systems or food processing systems, and others.~~

9 Another advantage is that the present invention provides methods for readily and
10 efficiently determining therapeutic resonant frequencies that take into account an appropriate
11 electromagnetic refractive index for ~~wave propagation velocity associated with~~ a surrounding
12 medium. In so doing, the present invention has the advantage of determining a more precise
13 therapeutic resonant frequency for the genomic system ~~target nucleic acid chain~~ in a particular
14 medium.

15 Still another advantage is that the present invention provides easier and more efficient
16 methods for determining resonant frequencies that significantly enhance the therapeutic benefit
17 and cost-effectiveness of currently existing electromagnetic, magnetic, plasma, audio, or other
18 frequency-emitting ~~frequency-capable emission~~ devices.

19 Another advantage over prior approaches to identifying resonant frequencies is that the
20 present invention provides the advantage of methods that utilize a simple biophysical ~~or~~
21 ~~biochemical~~ model for explaining and understanding why specific resonant frequencies are ~~can~~
22 ~~be~~ effective.

As will be realized by those of skill in the art, many different embodiments of methods for determining therapeutic resonant frequencies according to the present invention are possible. Additional uses, objects, advantages, and novel features of the invention are set forth in the detailed description that follows and will become more apparent to those skilled in the art upon examination of the following or by practice of the invention.

DETAILED DESCRIPTION OF THE INVENTION

The present invention comprises methods for determining resonant frequencies having therapeutic or beneficial uses in a variety of settings. In particular, the present invention includes methods for efficiently and accurately determining therapeutic resonant frequencies for specific complete genomes, or partial genomic materials. ~~and nucleic acid chains.~~ Methods of the present invention also comprise means for determining a more precise, and thus more therapeutic resonant frequency for the genomic system ~~target nucleic acid chain~~ in a particular medium by accounting for an appropriate electromagnetic refractive index ~~wave propagation velocity~~ for the surrounding medium.

Complete Genome

As described above, an object has a natural resonant frequency by the correlation of the length of the object with a wavelength that manifests into its surrounding medium. For example, the length of a DNA or RNA chain provides a wavelength ~~parameter~~ measurement that can be used to determine a resonant frequency. In embodiments of the present invention, the spacing of nucleotide base pairs in a DNA double helix is used in the mathematical process to determine frequency. The entire length of a ~~genome or other strand of DNA~~ piece of genomic material, is

1 determined by multiplying the number of base pairs or bases in the ~~genome or other strand of~~
2 ~~DNA genomic material~~ times the spacing length parameter between base pairs or bases.

3 It is known that base pair spacing in strands of DNA is not always consistent. Localized
4 areas contain “squeezing” or “spreading” of base pairs in various ways. In embodiments of the
5 methods of the present invention, the classic Watson-Crick model of base pair spacing is used.
6 The Watson-Crick model of base pair spacing is an average spacing over the entire length of the
7 DNA molecule. ~~Since lengths of target nucleic acid chains comprise some hundreds or~~
8 ~~thousands of base pairs (or nucleotides),~~ Use of an average base pair spacing allows for
9 accuracy sufficient to determine therapeutic resonant frequencies in accordance with the methods
10 of the present invention.

11 The B-helix is the most common in-vivo DNA form in bacterial and eukaryotic life
12 forms, and is used herein as illustration in the methods of the present invention. In the B-helix,
13 one complete turn of the helix spans a distance of 35.4 angstroms on its axis; and there are 10.4
14 base pairs in each helical turn. Therefore, the spacing of individual base pairs on the axis would
15 be 35.4 angstroms per turn divided by 10.4 base pairs per turn, which equals 3.403846 angstroms
16 spacing between each base pair. In scientific notation using SI units, the base pair spacing length
17 is expressed as 3.403846 e-10 meters. ~~because one angstroms equals 1 e-10 meters.~~ This use of
18 meters allows ~~The use of meters is required to compute the~~ conversion of the total length
19 ~~parameter of the DNA chain~~ (treated as wavelength) into a frequency.

20 By way of illustration using a pathogenic microorganism, the DNA genome of *Borrelia*
21 *burgdorferi* strain B31 contains 910,724 base pairs. To determine ~~wavelength~~ its length,
22 910,724 base pairs times the base pair spacing of 3.403846 e-10 meters = 3.09996 e-4 meters
23 total length of the genome. As described above, the length of an object can represent the object's

wavelength; in this case, the length of the *Borrelia* genome represents its a wavelength. ~~which can then be used for the frequency calculation.~~

To convert this wavelength to frequency, the following common physics relationship is used:

$$\text{velocity} / \text{wavelength} = \text{frequency} \quad (1)$$

If the DNA under consideration was in a medium of air, velocity would be the speed of electromagnetic radiation, or light, in air. For purposes of comparison, if *Borrelia burgdorferi* was in an air medium, according to methods of the present invention, the velocity of electromagnetic radiation emission through air (299,792,458 m/s) would be used in determining a therapeutic resonant frequency. Dividing this velocity by the *Borrelia burgdorferi* genome wavelength: (299,792,458 m/s / 3.09996 e-4 meters) = 9.6708492 e+11 Hz, ~~which would constitute a~~ the therapeutic resonant frequency for *Borrelia burgdorferi* in an air medium.

However, genomic nucleic acid material including that of *Borrelia burgdorferi*, generally ~~often~~ exists in a medium of living tissue. The velocity of electromagnetic radiation emission through a general in-vivo tissue medium is equal to the inverse of the square root of the product of the electrical permittivity and the magnetic permeability of the medium. The formula for velocity of electromagnetic radiation through a typical in-vivo tissue medium is given as:

$$\text{velocity} = 1 / \sqrt{(\epsilon \mu)} \quad (2)$$

where ϵ is the ~~unique~~ electrical permittivity of the medium, and μ is the magnetic permeability of the medium.

The magnetic permeability (μ) through in-vivo tissue ~~and most other biological substances~~ is known to be the same as that in air: 1.2566370614 e-6 henrys / meter ~~and therefore is not a unique parameter.~~ However, electrical permittivity in live body tissue ~~(and many other~~

materials) is not the same as for air. A representative value for electrical permittivity through in-vivo tissue is $71 \text{ e-12 farads / meter}$. Applying these figures to formula (2) above, the result is: $\text{velocity} = 1/\sqrt{[(71 \text{ e-12 F/m}) \times (1.2566370614 \text{ e}^{-6} \text{ H/m})]} = 105,868,288.9 \text{ meters per second}$, a representative velocity of electromagnetic radiation ~~emission~~ through in-vivo tissue.

Thus, in this method of the present invention, to obtain an in-vivo therapeutic resonant frequency of the *Borrelia burgdorferi* DNA genome having a ~~wavelength-associated parameter~~ length of $3.09996 \text{ e-4 meters}$, formula (1) above ($\text{velocity} / \text{wavelength} = \text{frequency}$) is then used: ~~to calculate a resonant frequency:~~ $105,868,288.9 \text{ meters per second} / 3.09996 \text{ e-4 meters} = 3.41515016 \text{ e+11 Hz}$.

Using the results of the above steps, a general refractive index of electromagnetic radiation ~~emission~~ through in-vivo tissue can be determined. A refractive index (n) is given by the ratio of the speed of light in a vacuum to the speed of light in the medium under consideration. This ratio is stated as:

$$n = \text{speed of light in a vacuum} / \text{speed of light in a medium.} \quad (3)$$

According to the steps given above, a refractive index of electromagnetic radiation through in-vivo tissue would be: $(299,792,458 \text{ m/s}) / (105,868,288.9 \text{ m/s}) = 2.831749$.

Then, by dividing a therapeutic frequency determined for a particular genomic material in an air medium by the refractive index for in-vivo tissue, a therapeutic resonant frequency for the genomic material in an in-vivo tissue medium is quickly determined. ~~An alternative method can be easily employed using this refractive index, to calculate a resonant frequency for a target nucleic acid chain in in-vivo tissue.~~ Following the example above, dividing the resonant frequency of *Borrelia* in air ($9.6708492 \text{ e+11 Hz}$) by the refractive index of electromagnetic

1 radiation emission through in-vivo tissue (2.831749), ~~gives will also give~~ the in-vivo resonant
2 frequency for the *Borrelia burgdorferi* genome (3.41515016 e+11 Hz).

3 The steps described above for the methods of the present invention can be adjusted to
4 correlate with any medium surrounding the genomic material ~~that may be surrounding the~~
5 ~~nucleic acid chain~~ under consideration, as long as an accurate electromagnetic velocity through
6 the medium is known or can be determined. ~~from its electrical permittivity or accurate refractive~~
7 ~~index characteristics, as described above.~~

8 The 3.41515016 e+11 Hz in-vivo therapeutic resonant frequency determined above for
9 the *Borrelia burgdorferi* genome ~~is a frequency that lies~~ appears in the infrared range of the
10 electromagnetic spectrum. In embodiments of the present invention, methods allow access to
11 corresponding resonant frequencies in the lower audio range. ~~lower radio or audio ranges of the~~
12 ~~electromagnetic spectrum.~~ For example, to determine an accurate resonant frequency in the
13 audio ~~electromagnetic~~ range corresponding to ~~the first original~~ a first therapeutic resonant
14 frequency, ~~as calculated above~~, the first resonant frequency is divided by the number 2, as many
15 times as necessary, to reach a frequency in the audio range. In musical terms, as described
16 above, frequencies that are related by a factor of 2, or a power thereof, are known as octaves. In
17 the example of the in-vivo *Borrelia burgdorferi* genome, a multi-octave shift to audio range can
18 be reached by dividing the first ~~original~~ therapeutic resonant frequency by 2^{29} , which gives a
19 corresponding second useful therapeutic resonant frequency of 636.12 Hz, which is in the audio
20 range. This process of dividing (or multiplying) any resonant frequency transposes it into a
21 different octave by ~~respectively~~ doubling (or halving) its wavelength in an exact and precise
22 manner, allowing a resonant correlation with the length parameter under consideration in a
23 specific medium. Thus, in the present invention, an octave-shifted translated therapeutic

1 resonant frequency will have a precise correlation with the first therapeutic ~~original~~ resonant
2 frequency. ~~Each of these frequencies will resonate with and amplify the other to provide~~
3 ~~enhanced beneficial effect.~~

4 In the example above, an in-vivo therapeutic resonant frequency of the *Borrelia*
5 *burgdorferi* genome is 3.41515016 e+11 Hz. Corresponding therapeutic useful resonant
6 frequencies in a different electromagnetic range, determined by dividing by appropriate powers
7 of 2, results in ~~produces~~ *Borrelia burgdorferi* in-vivo therapeutic resonant frequencies in the
8 audio range at: 636.12 Hz, 1272.24 Hz, 2544.5 Hz, 5088.9 Hz, etc.

9 As another illustration, if *Borrelia burgdorferi* were ~~theoretically~~ in a different medium
10 such as water at 40 degrees centigrade, according to methods of the present invention, the
11 velocity of EM radiation ~~electromagnetic emissions~~ through water at that temperature
12 (225,319,768 m/s) would be used in determining therapeutic resonant frequencies. Dividing this
13 velocity by the genome ~~genome-associated wavelength length: stated above:~~ (225,319,768 m/s) /
14 (3.09996 e-4 meters) = 7.2684734 e+11 Hz, which would be the therapeutic resonant frequency
15 of *Borrelia burgdorferi* DNA in ~~surrounded by~~ water at 40 degrees centigrade.

16 To determine corresponding therapeutic resonant frequencies in a different
17 electromagnetic frequency range, again in this instance the audio range, the resulting resonant
18 frequency ~~given~~ above is then divided by appropriate powers of 2. This gives therapeutic
19 resonant frequencies in the audio range for *Borrelia burgdorferi* in a 40-degree centigrade water
20 medium of: 676.9 Hz, 1353.9 Hz, 2707.7 Hz, 5415.4 Hz, etc.

21 In an alternative embodiment of the present invention, methods for determining
22 therapeutic resonant frequencies for a ~~nucleic acid chain~~ genomic material under consideration
23 use the ~~constitutes using a simple mathematical short cut method which eliminates almost all of~~

1 ~~the tedious numerical calculations described above. For example, to produce a useful resonant~~
2 ~~frequency in the audio range, the numerical constant 4,526,016.44 can be used as follows:~~
3 4,526,016.44 divided by the number of ~~nucleotides~~ base pairs or bases in a chain = frequency.
4 As such, ~~use of this particular method provides an efficient and simple means for determining a~~
5 ~~useful resonant frequency in the audio range. by ascertaining the number of base pairs or bases in~~
6 ~~the genomic material, and dividing that number into the aforementioned constant.~~ For example,
7 if there are 250 base pairs, or bases ~~nucleotides~~ in a ~~nucleic acid~~ DNA chain, 4,526,016.44
8 4,526,016.44 / 250 = 18,104.07 hertz. For 5,000 base pairs or bases ~~nucleotides~~ in a ~~nucleic acid~~
9 DNA chain, 4,526,016.44 4,526,016.44 / 5,000 = 905.20 hertz. For 22,000 base pairs or bases
10 ~~nucleotides~~ in a ~~nucleic acid~~ DNA chain, 4,526,016.44 4,526,016.44 / 22,000 = 205.73 hertz.

11
12
13 5 full paragraphs from the previously submitted specification should be deleted here.
14 They are located on pages 21-23 of that submission. The text of the first paragraph begins "This
15 short-cut method is derived...", and the text of the fifth paragraph ends with the number
16 "70,719.007".

17
18
19 As described above, in methods of the present invention, therapeutic additional ~~additional~~ resonant
20 frequencies are also ~~can be~~ determined for a ~~slightly~~ different electromagnetic range, for example
21 in ~~other areas of~~ the audio range, by dividing (or multiplying) by appropriate powers of 2. Using
22 the example of a 250-base pair DNA chain above, 18,104.07 Hz / 2 = 9,052.035 Hz. Repeated
23 division of the resulting frequency by a factor of 2, such that 9,052.035 Hz / 2 = 4526.017 Hz / 2

1 = 2263.008 Hz / 2 = 1131.504 Hz / 2 = 565.752 Hz, quickly ~~produces a useful frequency~~
2 determines frequencies in a the range capable of generation by typical frequency-emitting
3 devices. ~~numerous frequency-capable emission devices.~~ To further shorten the ~~An alternate and~~
4 ~~even faster method of performing this process, is by dividing 18,104.07 hz by 32, or 2⁵ (2 to the~~
5 ~~power of 5), which yields a frequency of 565.752 Hz. Multiplying or dividing by an appropriate~~
6 ~~factor of 2 (2, 4, 8, 16, 32, 64, 128, 526, etc.) will accurately convert~~ therapeutic resonant
7 ~~frequencies to a desired range for use in currently available frequency capable emission devices.~~
8 ~~Shifting or translating~~ frequencies by factors of 2 produces a frequency event that is ~~occurring at~~
9 an octave-related resonant frequency and wavelength.

10 As described above, many currently available frequency-emitting ~~frequency-capable~~
11 ~~emission~~ devices are not capable of producing therapeutic ~~able to accurately emit an original~~
12 ~~resonant frequencies~~ frequency in the infrared (~~or nearby~~) range, as that determined for the
13 *Borrelia burgdorferi* genome. To overcome such limitations, methods of the present invention
14 adjust resonant frequencies ~~upward or downward~~ (or upward) by dividing (or multiplying) by a
15 power of 2, until a frequency in the frequency-generating device's range of a device is achieved.
16 ~~frequency capability is reached.~~

17 Certain frequency ~~capable emission~~ devices emit not only a basic frequency (also referred
18 to as the “fundamental” frequency), but also many harmonics of that frequency. A “harmonic” is
19 defined as a positive integer multiple of the fundamental frequency. On this basis, in methods of
20 the present invention, additional ~~useful~~ frequencies can be determined and programmed into a
21 frequency-emitting ~~frequency-capable emission~~ device such that a harmonic of a frequency
22 corresponding to a first therapeutic resonant ~~a fundamental resonant frequency in any part of the~~
23 ~~spectrum, of associated with a target genomic material, nucleic acid chain,~~ would be emitted

1 along with the fundamental ~~resonant~~ frequency. Similar additional ~~useful~~ frequencies can be
2 determined by dividing the therapeutic resonant frequency by a positive integer, resulting in a
3 “subharmonic” frequency. Subharmonic frequencies corresponding related to a first therapeutic
4 ~~fundamental~~ resonant frequency of a target genomic material ~~nucleic acid chain~~ could also be
5 programmed into a frequency-emitting ~~frequency-capable emission~~ device, and be emitted along
6 with the fundamental ~~resonant~~ frequency. In this manner, a group of resonant frequencies
7 corresponding related to the first therapeutic ~~fundamental~~ resonant frequency can be emitted
8 simultaneously. As a result, effectiveness of a particular device can be enhanced. And as is well
9 ~~known to those skilled in the art, selection of certain waveforms offered by some frequency-~~
10 ~~capable emission devices, will also make possible automatic inclusion of various harmonies~~
11 ~~inherently present in the particular waveform chosen for the emission.~~

12 As an example, ~~To further demonstrate use of, for example, a subharmonically related~~
13 ~~frequency,~~ one in-vivo *Borrelia burgdorferi* therapeutic resonant frequency in an audio-range
14 octave is 636.12 Hz. When this therapeutic resonant frequency is divided by the positive integer
15 2, the resulting subharmonic frequency is 318.06 Hz. When this subharmonic frequency is
16 programmed into a harmonic-rich output device and emitted, the audio-range therapeutic
17 resonant frequency 636.12 Hz is emitted simultaneously, increasing the likelihood that a
18 therapeutic resonant frequency will impinge a target *Borrelia burgdorferi* genome. In like
19 manner, when dividing the audio-range therapeutic resonant frequency 636.12 Hz by the positive
20 integer 3, the resulting subharmonic frequency is 212.04 Hz. A harmonic-rich output device
21 programmed with this subharmonic frequency would also emit the 636.12 Hz therapeutic
22 resonant frequency, further increasing the likely efficacy of the treatment.

1 The in-vivo therapeutic resonant frequency determined in the audio range for the *Borrelia*
2 *burgdorferi* genome (636.12 Hz) is very close to a frequency (640 Hz) commonly used for Lyme
3 disease, which is caused by *Borrelia burgdorferi*. The accuracy of the methods of the present
4 invention may be ~~further~~ confirmed by comparing the resultant therapeutic resonant frequencies
5 produced by these methods, with many known ~~numerous previously used~~ and publicly available
6 therapeutic frequencies. ~~many of which are available for review at~~
7 ~~<http://www.electroherbalism.com/Bioelectronics/FrequenciesandAnecdotes/CAFL.htm>, and~~
8 ~~various other public websites.~~

9 In another example using a different pathogen, the Rubella measles RNA virus contains
10 9755 ~~nucleotides~~ bases in its entire genome. (9755 nucleotides) x (the ~~nucleotide~~ spacing of
11 3.403846 e-10 meters) = 3.32045 e-6 meters total length. This length is ~~then~~ used as the a
12 wavelength ~~for to influence~~ the Rubella viral genome. To obtain the in-vivo therapeutic resonant
13 frequency ~~for of~~ this wavelength, formula (1) above is again used: (105,868,288.9 meters per
14 second) / (3.32045 e-6 meters) = 3.188371724 e+13 Hz. ~~Subsequent octave adjustment~~ A
15 shifting of this near-infrared frequency to audio range by dividing by 2^{36} , gives a frequency of
16 463.97 Hz. A known therapeutic ~~previously used~~ frequency for the condition of Rubella measles
17 is 459 Hz, ~~which reveals~~ which is another close match to the therapeutic to a Rubella genome-
18 ~~related~~ resonant frequency determined by the methods of the present invention.

19 A number of favorable responses have been reported by individuals using previously
20 unknown therapeutic resonant frequencies determined by methods of the present invention. For
21 example, one person who often experienced severe outbreaks of herpes simplex virus used the
22 genome-related therapeutic resonant frequencies derived by the methods of the present invention
23 for several strains of herpes simplex viruses. This individual reported a much faster healing

process than what is usually experienced. Another example involves a person suffering from cancerous cervical warts. After use of previously unknown therapeutic resonant frequencies relating to the genome of a strain of papilloma virus derived by the methods of the present invention, this person reported disappearance of the warts. Still another example is a person infected with the chickenpox virus, who used a previously unavailable therapeutic audio-range resonant frequency derived by the methods of the present invention and associated with the varicella virus genome. This person reported rapid disappearance of blisters and symptoms associated with this disease.

In addition, in-vitro laboratory testing demonstrated that exposure of a ~~non-pathogenic~~ strain of *Escherichia coli* to a its genome-related resonant frequency produced a statistically significant reduction in the number of colonies in cultures.

~~Additional case results are presented in fuller detail at the end of this description.~~

Genes and Gene Sections

Methods of the present invention for determining therapeutic resonant frequencies as described above can also be applied to sections of DNA and/or RNA, as in genes, for example. Using genetic coding information, methods of the present invention for determining therapeutic resonant frequencies may also be utilized with other sub-components of genomic material, such as the coding associated with enzymes, immune factors, oncogenes, oncogenic growth factors, and other proteins.

In embodiments of the present invention, therapeutic resonant frequencies are determined using basic information about a protein, for example, how many amino acids are in the protein chain. Because an amino acid is always coded by three ~~nucleotides~~ bases in the messenger RNA, the number of ~~nucleotides~~ bases for use in determining resonant frequencies can be ascertained

1 by multiplying the number of amino acids in a protein chain by 3. For example, if there are 100
2 amino acids in a protein chain, there would be 300 ~~nucleotides~~ bases in the final messenger RNA
3 related to that protein. Thus, according to methods of the present invention, a therapeutic
4 resonant frequency can be easily determined: ~~with the previously mentioned shortcut method~~
5 ~~using a constant:~~ $4,526,016.44 / 300 \text{ ~~nucleotides~~ } = 15,086.72 \text{ Hz.}$
6 Using a factor of 2^5 to determine a corresponding therapeutic resonant frequency in a lower
7 octave within the acoustic range as described in the methods of the present invention above, the
8 resulting therapeutic resonant frequency would be: $15,086.72 \text{ Hz} / 32 = 471.46 \text{ Hz.}$ which is a
9 frequency that currently available frequency-emitting devices are capable of generating.

10 As an example, the int-1 mammary oncogene contains 4522 base pairs of DNA. A
11 therapeutic resonant frequency for this oncogene determined by the methods of the present
12 invention above is 2001.77 Hz. This therapeutic resonant frequency is very close to 2008 Hz, a
13 commonly used cancer-related frequency. Furthermore, the messenger RNA associated with the
14 final form of the transforming protein of the int-1 mammary oncogene contains 1112 ~~nucleotides~~
15 bases. A therapeutic resonant frequency for this transforming protein determined by the methods
16 of the present invention above is 2035.08 Hz, which is also in a range of cancer-related
17 frequencies currently in use.

18 As another example, the messenger RNA for the cancer-associated enzyme human
19 tyrosine kinase contains 3151 ~~nucleotides~~ bases. A therapeutic ~~audio-range~~ resonant frequency
20 for this enzyme's messenger RNA, as determined by the methods of the present invention above,
21 is 2872.7 Hz. This frequency is very close to the cancer-related frequency 2876 Hz, which,
22 along with its related octaves, ~~thereof~~ have been used throughout most of the twentieth century in
23 association with certain cancer therapy modalities.

1 Another example is a precursor gene for *Borrelia burgdorferi* outer surface protein A
2 (ospA), which contains 822 base pairs. A therapeutic resonant frequency for this ~~protein's~~
3 ~~messenger-RNA gene~~ determined by the methods of the present invention above, after being
4 factored by powers of 2 to the audible range, is 344.13 Hz. A previously known frequency
5 currently used for therapy related to lyme disease is 344 Hz, nearly an exact match.

6 As can be seen, therapeutic resonant frequencies for genes, gene sections, and constituent
7 components of genomic material ~~and those for the precursor nucleotide chains of enzymes,~~
8 ~~proteins, and the like,~~ can be determined more readily and efficiently by methods of the present
9 invention than for example, by trial and error.

10 Favorable responses have ~~also~~ been reported ~~following~~ from the use of previously
11 unavailable therapeutic resonant frequencies determined by methods of the present invention,
12 relating to genes, components of genes, and/or messenger RNA coding associated with certain
13 proteins. For example, an individual diagnosed with lung cancer used therapeutic resonant
14 frequencies related to certain ~~cancer~~ growth factors and the K-ras oncogene, which is associated
15 with his type of tumor. It is reported that this individual experienced eradication of lung tumor
16 material. Another example is a student experiencing symptoms of both lyme disease and
17 ehrlichiosis, who was unable to attend school for a year and half due to the severity of
18 symptoms. The student used previously unavailable therapeutic resonant frequencies as
19 determined by methods of the present invention, for certain membrane and antigenic proteins
20 associated with the organism *Ehrlichia chaffeensis*. Within two weeks of beginning therapy with
21 those therapeutic resonant frequencies, this student was well enough to return to school.

Two full paragraphs should be removed at this location from the previously-submitted specification. They were located on page 30, lines 4-11, beginning with the text “There are numerous public internet...” and finishing with “repeatability of results”.

Case Results

8 examples of case results should be deleted from the previously-submitted specification, located on pages 30-35 of that submission.

While the present invention has been described with reference to several specific embodiments, those skilled in the art will be able to make various modifications to the described embodiments, for instance, by factoring therapeutic octave-adjusting resonant frequencies to electromagnetic ranges to other than audible ranges, ~~the audio range~~, and by adjusting for various media, without departing from the spirit and scope of the invention. It is therefore to be understood that within the scope of the appended claims the invention may be practiced other than as specifically described herein.

ABSTRACT

Methods are provided for readily and efficiently determining resonant frequencies that can be used therapeutically or beneficially, for ~~stimulation or~~ debilitation of specific types of genomic materials, including DNA and/or RNA, genes, and ~~or~~ gene sections. The methods can be used in a variety of circumstances ~~and settings~~, ~~The methods are intended to influence~~ ~~nucleic acid chains, including those~~ related to various ~~pathogenic~~ human and animal diseases and conditions, ~~agriculture-related diseases, pathogen-contamination of water systems or food~~

1 ~~processing systems, and others.~~ Methods allow determination of therapeutic resonant
2 frequencies ~~associated with nucleic acids,~~ for use in various media having different refractivities.
3 ~~which may be present in a variety of surrounding media that may have velocities of propagation~~
4 ~~of electromagnetic waves different from that of air.~~ Therapeutic or beneficial Useful resonance
5 frequencies thus determined are adapted for use with currently available frequency-emitting
6 ~~frequency-capable emission~~ devices by ~~translating~~ shifting resonant frequencies to
7 electromagnetic ranges capable of generation by such devices.

8
9